



## **CrossTalk opposing view**

### **lack of evidence supporting an astrocyte-to-neuron lactate shuttle coupling neuronal activity to glucose utilisation in the brain**

Bak, Lasse K; Walls, Anne B

*Published in:*  
The Journal of Physiology

*DOI:*  
[10.1113/JP274945](https://doi.org/10.1113/JP274945)

*Publication date:*  
2018

*Document version*  
Publisher's PDF, also known as Version of record

*Document license:*  
[Unspecified](#)

*Citation for published version (APA):*  
Bak, L. K., & Walls, A. B. (2018). CrossTalk opposing view: lack of evidence supporting an astrocyte-to-neuron lactate shuttle coupling neuronal activity to glucose utilisation in the brain. *The Journal of Physiology*, 596(3), 351-353. <https://doi.org/10.1113/JP274945>

## CROSSTALK

### CrossTalk opposing view: lack of evidence supporting an astrocyte-to-neuron lactate shuttle coupling neuronal activity to glucose utilisation in the brain

Lasse K. Bak  and Anne B. Walls

Department of Drug Design and Pharmacology, Faculty of Health and Medical Sciences, University of Copenhagen, 2 Universitetsparken, 2100 Copenhagen, Denmark

Email: laba@sund.ku.dk

Edited by: Francisco Sepúlveda & Ian Forsythe

In 1993, Dringen *et al.* concluded that 'glycogen in astrocytes can be considered as a store for lactate rather than for glucose', and suggested that lactate derived from the breakdown of glycogen in astrocytes may serve the energetic needs of neighbouring cells. The following year, Pellerin and Magistretti (1994) published their now famed astrocyte-to-neuron lactate shuttle hypothesis in which the transfer of lactate from astrocytes to neurons, in this case derived from extracellular glucose rather than glycogen, is coupled to uptake of neurotransmitter glutamate (i.e. neuronal activity). According to this hypothesis, glycolysis and lactate production are astrocytic phenomena while oxidative metabolism of lactate takes place in neurons. The astrocyte-to-neuron lactate shuttle hypothesis as proposed by Pellerin and Magistretti (1994) has gained widespread acceptance, and its popularity is not surprising due to its conceptually simple and compelling idea of an activity-based coupling between neuronal synaptic activity and astrocyte metabolism. We will argue that the biochemical and physiological

evidence for the existence of a unidirectional flow of lactate from astrocytes to neurons, as proposed, is lacking. However, before we get to that, let us briefly explore why this subject is even interesting to physiologists.

#### Why is this issue worth a CrossTalk debate?

First of all, the extensive acceptance of the astrocyte-to-neuron lactate shuttle means that many researchers use this hypothesis as a master template on which they interpret their data, thus ignoring alternative explanations and hence creating a bias in the literature. Besides this, and of course the pure scientific desire to know how the brain operates, the cellular site of glucose metabolism in the brain is important for interpreting fluorodeoxyglucose (FDG) positron emission tomography (PET) results, a method extensively used for both research and diagnostic purposes. In the following, we will focus on two key issues that we believe severely contest the existence of a lactate shuttle from astrocytes to neurons, as proposed: (1) neurons express glucose transporters and metabolise glucose in an activity-dependent manner; and (2) the distinct cellular isoform expression of lactate transporters and lactate dehydrogenase, the enzyme forming lactate from pyruvate, cannot be employed as an argument for a directional flow of lactate.

#### Neurons metabolise glucose in an activity-dependent manner suggesting that glucose is an important neuronal energy substrate during activation

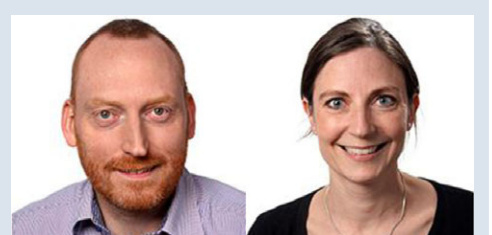
In support of the lactate shuttle hypothesis, it has been proposed that neurons do not metabolise glucose in an activity-dependent manner and that lactate is their preferred substrate (e.g. Bouzier-Sore *et al.* 2003). Contesting this view, we know that neurons express transport systems for glucose both *in*

*vitro* and *in situ* (Simpson *et al.* 2007), and others and we have repeatedly shown that both cultured neurons and synaptosomes (an *ex vivo* preparation of presynaptic neuronal terminals typically obtained from rodent brain) avidly take up and metabolise glucose in an activity-dependent manner (e.g. Bak *et al.* 2006; Patel *et al.* 2014). Further, Lundgaard *et al.* (2015) and Diaz-Garcia *et al.* (2017), employing a near-infrared 2-deoxyglucose probe and redox biosensors, respectively, showed that glucose is metabolised by neurons in an activity-dependent manner *in situ* in awake mice. As discussed further below there are no good biochemical reasons why neurons should primarily consume lactate during activation; indeed, neurons *in vitro* are, not surprisingly, able to produce lactate upon activation and thus neurons may contribute to the extracellular surge in lactate associated with brain activation (Prichard *et al.* 1991; Hu & Wilson, 1997; Bak *et al.* 2009; Contreras & Satrustegui, 2009). To be fair, it is important to note that neurons do metabolise lactate if present (Bak *et al.* 2009), and in the words of a long-time opponent of the lactate shuttle hypothesis, Gerry Dienel, lactate is an 'opportunistic' substrate that, if present, indeed will serve to support energy metabolism (Dienel, 2012).

#### Distinct isoform expression of lactate transporters and lactate dehydrogenase in neurons and astrocytes does not predict directionality of any shuttling of lactate

Monocarboxylate transport (MCT) systems, facilitative transporters allowing lactate or pyruvate to cross the plasma membrane, are present on both neurons and astrocytes and they differ in their kinetic profiles, i.e. their transport capacity and binding affinities for lactate (Simpson *et al.* 2007). These differences have been

**Lasse K. Bak** is an associate professor at the Department of Drug Design and Pharmacology at University of Copenhagen, Denmark. His research focuses on understanding compartmentalised cAMP and Ca<sup>2+</sup> signals, and signalling-metabolism coupling in brain cells in search of novel drug targets for brain pathologies such as dementias and epilepsy. **Anne B. Walls** is an associate professor at the Department of Drug Design and Pharmacology at University of Copenhagen, Denmark. Her research focuses on understanding energy and amino acid metabolism in brain cells and its coupling to cerebral activity in pathologies such as hepatic encephalopathy and epilepsy.



employed as arguments for a unidirectional flow of lactate from astrocytes to neurons (e.g. Bittar *et al.* 1996). However, regardless of the kinetic parameters of a facilitative transport system, the flow of substrate in either direction is governed by the prevailing intra- to extracellular concentration gradient, i.e. in this case the production vs. disappearance or consumption of lactate. Thus, an extensive, activity-dependent astrocyte-to-neuron gradient of lactate needs to be established for the lactate shuttle to work as suggested. Mächler *et al.* (2016) recently investigated this and we will get to that shortly. First, the preferential synthesis of lactate in astrocytes and consumption in neurons have been argued to be possible due to distinct cellular expression of isozymes of lactate dehydrogenase (LDH) having dissimilar kinetic parameters, e.g. in terms of binding constants for lactate (e.g. Laughton *et al.* 2000). However, regardless of their kinetic parameters, enzymes influence the speed at which the thermodynamic equilibrium is obtained but do not change the equilibrium of a chemical reaction. Thus, a distinct cellular distribution of LDH isozymes with different kinetic parameters does not predict which cells are producing and which are consuming lactate (please see Bak & Schousboe, 2017 for a detailed discussion). Further, Quistorff & Grunnet (2011a, b) argue that the differences in kinetic parameters determined for LDH at room temperature are not present at body temperature. Thus, the distinct kinetic parameters of LDH employed as an argument in favour of the shuttle may not be real. So, how can an extensive lactate gradient be formed? The only way that is possible is if astrocytes are relentlessly outpacing neurons in terms of glycolytic flux and lactate production during activation. As alluded to above, Mächler *et al.* have investigated if there is such a lactate gradient between astrocytes and neurons. Employing anaesthetised mice expressing a lactate biosensor specifically in neurons or astrocytes they show that both neurons and astrocytes take up lactate when present in the blood in excessive amounts consistent with the concept of lactate being an 'opportunistic' substrate. By measuring the rate of biosensor saturation in the presence of ammonium chloride to inhibit mitochondrial ATP production, and thus boost glycolysis and lactate production, they estimate that neurons have a lower baseline

level of lactate than do astrocytes. While this an interesting observation, it does not tell us if astrocytes outpace neurons in lactate production during activation.

### Final thoughts

In our minds, the current literature and the biochemical design of neurons and astrocytes are largely consistent with a situation according to which both neurons and astrocytes contribute to the surge in extracellular lactate during brain activation; either cell type may then consume lactate when available or the lactate may be dispersed and metabolised elsewhere or even leave the brain (Madsen *et al.* 1999; Hertz *et al.* 2014; Satrustegui & Bak, 2015). The cellular location and timing of lactate synthesis and consumption in the brain in health and disease largely remains an open question that deserves to be investigated with an open mind.

### Call for comments

Readers are invited to give their views on this and the accompanying CrossTalk articles in this issue by submitting a brief (250 word) comment. Comments may be submitted up to 6 weeks after publication of the article, at which point the discussion will close and the CrossTalk authors will be invited to submit a 'LastWord'. Please email your comment, including a title and a declaration of interest, to [jphysiol@physoc.org](mailto:jphysiol@physoc.org). Comments will be moderated and accepted comments will be published online only as 'supporting information' to the original debate articles once discussion has closed.

### References

- Bak LK & Schousboe A (2017). Misconceptions regarding basic thermodynamics and enzyme kinetics have led to erroneous conclusions regarding the metabolic importance of lactate dehydrogenase isoenzyme expression. *J Neurosci Res* **95**, 2098–2102.
- Bak LK, Schousboe A, Sonnewald U & Waagepetersen HS (2006). Glucose is necessary to maintain neurotransmitter homeostasis during synaptic activity in cultured glutamatergic neurons. *J Cereb Blood Flow Metab* **26**, 1285–1297.
- Bak LK, Walls AB, Schousboe A, Ring A, Sonnewald U & Waagepetersen HS (2009). Neuronal glucose but not lactate utilization is positively correlated with NMDA-induced neurotransmission and fluctuations in cytosolic  $\text{Ca}^{2+}$  levels. *J Neurochem* **109**, 87–93.

- Bittar PG, Charnay Y, Pellerin L, Bouras C & Magistretti PJ (1996). Selective distribution of lactate dehydrogenase isoenzymes in neurons and astrocytes of human brain. *J Cereb Blood Flow Metab* **16**, 1079–1089.
- Bouzier-Sore AK, Voisin P, Canioni P, Magistretti PJ & Pellerin L (2003). Lactate is a preferential oxidative energy substrate over glucose for neurons in culture. *J Cereb Blood Flow Metab* **23**, 1298–1306.
- Contreras L & Satrustegui J (2009). Calcium signaling in brain mitochondria: interplay of malate aspartate NADH shuttle and calcium uniporter/mitochondrial dehydrogenase pathways. *J Biol Chem* **284**, 7091–7099.
- Diaz-Garcia CM, Mongeon R, Lahmann C, Koveal D, Zucker H & Yellen G (2017). Neuronal stimulation triggers neuronal glycolysis and not lactate uptake. *Cell Metab* **26**, 361–374.e4.
- Dienel GA (2012). Brain lactate metabolism: the discoveries and the controversies. *J Cereb Blood Flow Metab* **32**, 1107–1138.
- Dringen R, Gebhardt R & Hamprecht B (1993). Glycogen in astrocytes: possible function as lactate supply for neighboring cells. *Brain Res* **623**, 208–214.
- Hertz L, Gibbs ME & Dienel GA (2014). Fluxes of lactate into, from, and among gap junction-coupled astrocytes and their interaction with noradrenergic. *Front Neurosci* **8**, 261.
- Hu Y & Wilson GS (1997). A temporary local energy pool coupled to neuronal activity: fluctuations of extracellular lactate levels in rat brain monitored with rapid-response enzyme-based sensor. *J Neurochem* **69**, 1484–1490.
- Laughton JD, Charnay Y, Belloir B, Pellerin L, Magistretti PJ & Bouras C (2000). Differential messenger RNA distribution of lactate dehydrogenase LDH-1 and LDH-5 isoforms in the rat brain. *Neuroscience* **96**, 619–625.
- Lundgaard I, Li B, Xie L, Kang H, Sanggaard S, Haswell JD, Sun W, Goldman S, Blekot S, Nielsen M, Takano T, Deane R & Nedergaard M (2015). Direct neuronal glucose uptake heralds activity-dependent increases in cerebral metabolism. *Nat Commun* **6**, 6807.
- Mächler P, Wyss MT, Elsayed M, Stobart J, Gutierrez R, von Faber-Castell A, Kaelin V, Zuend M, San Martín A, Romero-Gómez I, Baeza-Lehnert F, Lengacher S, Schneider BL, Aebischer P, Magistretti PJ, Barros LF & Weber B (2016). In vivo evidence for a lactate gradient from astrocytes to neurons. *Cell Metab* **23**, 94–102.
- Madsen PL, Cruz NF, Sokoloff L & Dienel GA (1999). Cerebral oxygen/glucose ratio is low during sensory stimulation and rises above normal during recovery: excess glucose consumption during stimulation is not accounted for by lactate efflux from or accumulation in brain tissue. *J Cereb Blood Flow Metab* **19**, 393–400.

- Patel AB, Lai JC, Chowdhury GM, Hyder F, Rothman DL, Shulman RG & Behar KL (2014). Direct evidence for activity-dependent glucose phosphorylation in neurons with implications for the astrocyte-to-neuron lactate shuttle. *Proc Natl Acad Sci USA* **111**, 5385–5390.
- Pellerin L & Magistretti PJ (1994). Glutamate uptake into astrocytes stimulates aerobic glycolysis: a mechanism coupling neuronal activity to glucose utilization. *Proc Natl Acad Sci USA* **91**, 10625–10629.
- Prichard J, Rothman D, Novotny E, Petroff O, Kuwabara T, Avison M, Howseman A, Hanstock C & Shulman R (1991). Lactate rise detected by  $^1\text{H}$  NMR in human visual cortex during physiologic stimulation. *Proc Natl Acad Sci USA* **88**, 5829–5831.
- Quistorff B & Grunnet N (2011*a*). High brain lactate is not caused by a shift in the lactate dehydrogenase A/B ratio. *Proc Natl Acad Sci USA* **108**, E21; author reply E22.
- Quistorff B & Grunnet N (2011*b*). The isoenzyme pattern of LDH does not play a physiological role; except perhaps during fast transitions in energy metabolism. *Aging* **3**, 457–460.
- Satrustegui J & Bak LK (2015). Fluctuations in cytosolic calcium regulate the neuronal malate-aspartate NADH shuttle: implications for neuronal energy metabolism. *Neurochem Res* **40**, 2425–2430.
- Simpson IA, Carruthers A & Vannucci SJ (2007). Supply and demand in cerebral energy metabolism: the role of nutrient transporters. *J Cereb Blood Flow Metab* **27**, 1766–1791.

### Additional information

#### Competing interests

None declared.

#### Author contributions

Both authors have contributed to the conception or design of the work and drafting the work

or revising it critically for important intellectual content. Both authors have approved the final version of the manuscript and agree to be accountable for all aspects of the work. All persons designated as authors qualify for authorship, and all those who qualify for authorship are listed.

#### Funding

The Lundbeck Foundation is cordially acknowledged for funding A.B.W. (grant. no. R165-2013-15334).

### Supporting information

The following supporting information is available in the online version of this article.

Comments.

Last words by Barros & Weber.

Last words by Bak & Walls.